

Effect of Irradiation on Amino Acid *in vitro* Incorporation in the Course of Liver Regeneration

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Protein biosynthesis, irradiation, liver regeneration

The *in vitro* incorporation of ^{14}C -leucine by rat liver microsomes is studied under four different physiological states: 1. Normal liver of adult rats (NL), 2. irradiated (4 000 rads) liver (IL), 3. liver after partial hepatectomy (RL) and 4. regenerating liver in preirradiated rats (IRL) 2, 5, 12, and 24 hours after treatment.

Both irradiation and partial hepatectomy, when applied separately, stimulate the endogenous activity of microsomes, whereas preirradiation abolishes the stimulatory effect in the early hours after partial hepatectomy.

The results are in agreement with the concept that irradiation stimulates the operating synthetic programme of the cell and inhibits cellular reprogramming.

It has been suggested¹ that irradiation does not suppress directly the operating synthetic programme of the cell but prevents the process of cellular reprogramming. Studies on the radiosensitivity of RNA biosynthesis in regenerating liver² have supported this view by showing that irradiation enhances the synthesis of ribosomal RNA (rRNA) and DNA-like RNA (dRNA) in normal liver but inhibits the synthesis of dRNA in regenerating liver. The purpose of the present study was to check this hypothesis on the level of protein biosynthesis using rat liver in four different physiological states: 1. Normal liver of adult rats (NL), 2. liver of adult irradiated rats (IL), 3. liver after partial hepatectomy (RL), and 4. regenerating liver in preirradiated rats (IRL).

The results presented in this paper show that both irradiation and partial hepatectomy, when applied separately, stimulate the endogenous activity of microsomes, whereas preirradiation abolishes the stimulatory effect in the early hours after partial hepatectomy.

Methods and Materials

Treatment of experimental animals

The experiments were performed on 200 male albino rats (160–180 g body wt.), fasted 18 hours before killing.

Abbreviations: Pyruvate kinase — E. C. 2.7.1.40.

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The animals were irradiated with 4 000 rads of γ -rays by a Co^{60} unit under conditions ensuring homogeneity of the field controlled by a ferro-sulphate dosimeter. The dose rate was 159 rads/min. Liver regeneration (compensatory hypertrophy) was induced by removing two thirds of the liver by the method of HIGGINS and ANDERSON³, 30 min after irradiation.

Preparation of purified supernatant (S-100) and microsomes

The animals were killed by decapitation under light ether anaesthesia. The liver was rinsed and chilled in ice-cold medium A (0.05 M Tris/HCl, pH 7.6, 0.025 M KCl, 0.005 M MgCl_2 , 0.25 M Sucrose), minced with scissors, then homogenized in 2.5 volumes of medium A by means of a Potter-Elvehjem glass-Teflon homogenizer (two up-and-down strokes, 1800 rpm, clearance - 0.2 mm). The homogenate was centrifuged 15 min at $10\,000 \times g$ to remove cell debris, nuclei and mitochondria. The upper two thirds of the supernatant were centrifuged 60 min at $105\,000 \times g$ to sediment microsomes. The clear part of the postmicrosomal supernatant was dialysed overnight against 50 volumes of medium A with 0.005 M β -mercaptoethanol, frozen in small portions and stored at -20°C (S-100). For isolation of microsomes the livers were homogenized with 4 volumes of medium A and the preparation was performed as described above. The microsomal pellets were suspended in medium A by gentle manual homogenization and used immediately.

Estimation of RNA content of microsomes

RNA was determined spectrophotometrically with a correction for ferritin⁴.



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Method of incubation

The endogenous activity of microsomes was determined by cell-free incorporation of ^{14}C -leucine. The incubation mixture contained in one ml the following components in micromoles: MgCl_2 - 10, Tris/HCl, pH 7.6 - 30, β -mercaptoethanol - 2, ATP - 1, GTP - 0.4, PEP - 10, PEP kinase (E. C. 2.7.1.40) - 30 μg , ^{14}C -leucine (85mCi/mM - 1 μCi , mixture of all amino acids without leucine) - 0.01 μmole of each, sRNA and soluble enzymes added as S-100 from normal liver (about 2 mg of protein), microsomes - about 15 OD_{260} units. Incubation was at 37°C for the time specified.

After incubation the protein was precipitated and washed as described by CAMPBELL *et al.*⁵. The dried precipitate was dissolved in 85 % formic acid, transferred to aluminum planchets, dried at room temperature and counted in a gas-flow counter (VA-Z-530, Vakutronik).

Results and Discussion

In order to set up appropriate conditions for a precise comparative study the kinetics of the incorporation were examined (Fig. 1 A, B). As seen the

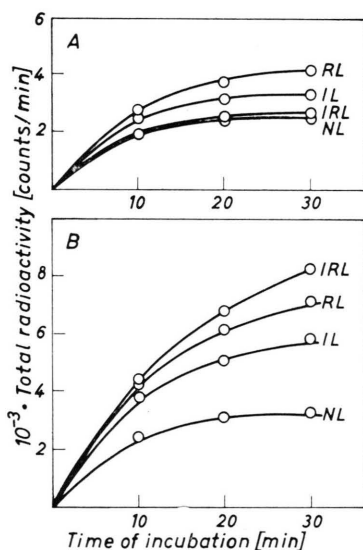


Fig. 1. Kinetics of ^{14}C -leucine incorporation by microsomes from normal liver (NL), irradiated liver (IL), regenerating liver (RL) and regenerating liver in preirradiated rats (IRL). A: 5 hours after treatment; B: 24 hours after treatment. For details see Methods.

incorporation is linear only for time intervals shorter than 10 min and a saturation level for the controls is reached in about 20 min, whereas microsomes from the three experimental groups continue to incorporate ^{14}C -leucine. This effect of increased longevity of

incorporation has been described for regenerating liver only by some authors^{6, 7} but it has not been found by others⁸. The effect is more expressed in the later hours following treatment (Fig. 1B). On the basis of these results a 20 min incubation time giving well pronounced differences between microsomes of normal and treated animals was used in all experiments.

The ^{14}C -leucine incorporation *in vitro* was studied 2, 5, 12, and 24 hours after treatment. As shown in Fig. 2 no stimulatory effect is observed at the 2nd

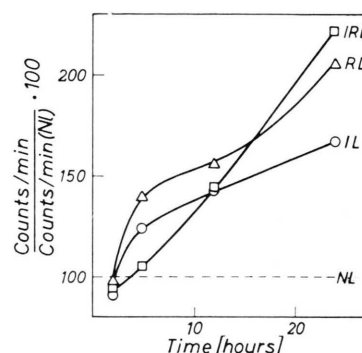


Fig. 2. ^{14}C -leucine incorporation by microsomes from irradiated liver (IL), regenerating liver (RL) and regenerating liver in preirradiated rats (IRL) isolated at various times after treatment. Incubation - 20 min at 37°C . Results expressed as per cent of the controls. For details see Methods.

hour. In fact, in all cases there is a slight decrease, which is consistent with the predominance of catabolic processes during this period after partial hepatectomy⁹. However, this decrease is not significant and needs further proof. Beginning from the 5th hour, both irradiation and partial hepatectomy, when applied separately, lead to a significant rise in the incorporating activity of microsomes, which is most pronounced at the 24th hour.

Different results are obtained with the combination of the two treatments. In this case the stimulatory effect at the 5th hour is strongly suppressed. At the 12th and the 24th hour an increased incorporation is observed again, although there is no additivity in the effects of the two treatments.

Our results concerning irradiated and partially hepatectomized animals are in good agreement with literature data showing that both partial hepatectomy^{6,8,10} and irradiation^{11,12} greatly stimulate the incorporating ability of microsomes. The absence of such a stimulatory effect in the early hours after partial hepatectomy in preirradiated animals has not

been observed earlier and is of special interest. This finding suggests that different mechanisms are involved in these two cases. Most probably irradiation increases the endogenous activity of microsomes of intact liver by stimulating the operating synthetic programme of the hepatocytes. This view is favoured also by the fact that there is an enhancement of the synthesis of RNA after irradiation^{2, 11, 12, 13}. Some data indicate that this effect is not mediated through the adrenal system^{14, 15, 16}.

On the other hand, in the case of regenerating liver the increased endogenous activity of microsomes is obviously connected with the process of cellular reprogramming, leading to the synthesis of new RNA species and new proteins^{2, 17, 18}.

The lack of stimulation of ¹⁴C-leucine incorporation at the early stages of regeneration in preirradiated rats most probably implies that irradiation affects the process of cellular reprogramming inhibiting in this way the normally observed increased incorporation after partial hepatectomy.

That irradiation does not suppress the operating synthetic programme of the cell but rather the cellular reprogramming is further supported by the re-

sults obtained on the level of RNA biosynthesis. It has been found² that irradiation enhances the synthesis of rRNA and dRNA in normal liver but inhibits the synthesis of dRNA in regenerating liver, without affecting the increased synthesis of rRNA in the latter case.

It is worthy to mention that at later hours of liver regeneration in preirradiated rats the endogenous activity of microsomes is strongly stimulated again (see Fig. 2). At present it is difficult to explain the mechanisms of this late increased incorporation. It should be pointed out, however, that it is well correlated with the increased synthesis of ribosomal RNA², the increased amount of RNA per liver cell (our unpublished data) and the increased weight of regenerating liver¹⁹ in preirradiated rats.

In conclusion, the results presented are in favour of the concept that irradiation stimulates the operating synthetic programme of the cell but inhibits cellular reprogramming.

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¹ G. G. MARKOV, Adv. Biochem. Biophysics (Bulgarian) **1**, 191 [1965].

² G. G. MARKOV, G. N. DESSEV, G. C. RUSSEV, and R. G. TSANEV, 7th Meeting FEBS, Abstracts p. 332, Varna 1971.

³ G. M. HIGGINS and R. M. ANDERSON, Arch. Path. **12**, 186 [1931].

⁴ S. H. WILSON and M. B. HOAGLAND, Proc. nat. Acad. Sci. USA **54**, 600 [1965].

⁵ P. N. CAMPBELL, G. SERCK-HANSEN, and E. LOWE, Biochem. J. **97**, 422 [1965].

⁶ M. B. HOAGLAND, O. A. SCORNIK, L. C. PFEFFERKORN, Proc. nat. Acad. Sci. USA **51**, 1184 [1964].

⁷ D. J. MCCORQUODALE, E. G. VEACH, and G. C. MUELLER, Biochim. biophysica Acta [Amsterdam] **46**, 335 [1961].

⁸ P. N. CAMPBELL, E. LOWE, and G. SERCK-HANSEN, Biochem. J. **103**, 280 [1967].

⁹ R. G. TSANEV and G. G. MARKOV, Folia Histochem. Cytochem. **2**, 233 [1964].

¹⁰ A. VON DER DECKEN and T. HULTIN, Exp. Cell Res. **14**, 88 [1958].

¹¹ E. J. HIDVEGI, J. HOLLAND, E. BÖLÖNI, P. LÓNAI, F. ANTONI, and V. VÁRTÉRESZ, Biochem. J. **109**, 495 [1968].

¹² E. J. HIDVEGI, E. BÖLÖNI, J. HOLLAND, F. ANTONI, and V. VÁRTÉRESZ, Biochem. J. **116**, 503 [1970].

¹³ V. L. MANTIEVA, E. A. RAPOPORT, S. G. TULKES, and J. B. ZBARSKY, Vopr. med. Khimii **12**, 407 [1966].

¹⁴ S. T. TAKETA, H. A. LEON, B. L. CASTLE, W. H. HOWARD, and S. DALIGKON, Rad. Res. **25**, 247 [1965].

¹⁵ P. CAMMARANO, S. PONS, G. CHINALI, and S. GAETANI, Rad. Res. **39**, 289, [1969].

¹⁶ P. G. POPOV, L. I. VALEVA-DIMITROVA, and A. A. HADJIOLOV, Z. Naturforsch. **26b**, 1282 [1971].

¹⁷ N. L. BUCHER, Int. Rev. Cytol. **15**, 245 [1963].

¹⁸ R. B. CHURCH and B. J. MCCARTHY, J. molecular Biol. **23**, 459 [1967].

¹⁹ G. G. MARKOV, P. PETROV, G. N. DESSEV, and R. G. TSANEV, Bull. Biochem. Res. Lab. **4**, 215 [1971].